

TERPENOIDS—LXXII¹

CHEMICAL STUDIES OF MARINE INVERTEBRATES—XXVI² $\Delta^{9(12)}$ -CAPNELLENE-3 β ,8 β ,10 α ,14-TETROL, A NOVEL POLYOXYGENATED SESQUITERPENE FROM THE ALCYONARIAN *CAPNELLA IMBRICATA*

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Abstract—The structure determination of the new sesquiterpene, $\Delta^{9(12)}$ -capnellene-3 β ,8 β ,10 α ,14-tetrol **1**, is described.

In previous publications,^{1,3} we described the isolation and structure determination of four representatives of the hitherto unknown capnellane skeleton from the soft coral *Capnella imbricata*. We wish now to report the structure determination of a fifth highly oxygenated member of this class of sesquiterpenes, the tetrol **1** so far only found in samples collected at Leti, Province of Maluku, Indonesia, and absent in *Capnella* collected from many other localities.¹

The NMR spectrum (1:1 mixture of *d*₅-pyridine and D₂O) of tetrol **1** (C₁₅H₂₄O₄; IR 3400 (OH), 905 cm⁻¹ (C=CH₂)) suggests the presence of two tertiary methyl groups (s, 3H each at 1.42 and 1.68), one primary (dd, 2H at 3.82), one allylic secondary (m, 1H at 5.33), one secondary non-allylic hydroxyl groups (m, 1H at 4.6) and one vinylic methylene (m, 2H at 5.40).

Acetylation of **1** furnished the triacetate **2** whose IR spectrum (ν_{OH} at 3500 cm⁻¹) implies the presence of a tertiary alcohol. Its NMR is similar to that of $\Delta^{9(12)}$ -capnellene-3 β ,8 β ,10 α -triol diacetate **3**¹ but shows clearly that hydroxylation of one of the tertiary methyl groups of **3** had occurred (AB system centered at 4.1 ppm). This was further substantiated by the ¹³C NMR spectrum of **1** wherein one of the three tertiary methyl group signals of the capnellane skeleton is lacking and replaced by one triplet at 73.84 ppm. Oxidation of **1** with manganese dioxide furnished the oily α,β -unsaturated ketone **4**, thus confirming the presence of an allylic secondary alcohol. The capnellane skeleton of **1** was established by the following reaction sequence: Li/NH₃ reduction of **4** afforded the keto-triol **5** which, without preliminary purification, was dehydrated to the α,β -unsaturated

ketone **6**. Li/NH₃ reduction of **6** yielded the saturated keto-diol **7** (IR 1720 cm⁻¹) which upon tosylation (*p*-TsCl/pyridine) to the ditosylate **8** and immediate reduction with LiAlH₄ in THF under reflux, led to capnellane-8 β -ol **9**. Jones oxidation of **9** yielded the monoketone **10** identical (IR, GC and MS) with the ketone derived from the previously described $\Delta^{9(12)}$ -capnellene-8 β ,10 α -diol **11**.¹ This established the nature of the skeleton of **1** and also the position of unsaturation at 9(12), a secondary hydroxyl at C-8 and a tertiary hydroxyl at C-10.

There remained only the location of the secondary non-allylic and primary alcohols. The primary hydroxyl function in **1** could only be attached to carbon atoms C-13, C-14 or C-15. The presence of a primary hydroxyl group at either C-14 or C-15 rather than C-13 was indicated as follows: Jones oxidation of **7** afforded the diketo-acid **12** (M⁺ 264) which could not be decarboxylated under acidic conditions (*p*-toluenesulfonic acid; 2N HCl) thus excluding C-13 as the site of the primary hydroxyl group. This conclusion could be further substantiated by Fetizon oxidation of **7** to the diketo-aldehyde **13** (M⁺ 246) which could not be de-formylated under either acidic or basic conditions.

Moreover, the mass spectra of **7**, **12**, **14** and **13** displayed peaks at *m/e* 101 (C₃H₇O₂, 13%); 114 (C₃H₆O, 70%); 128 (C₄H₈O, 100%) and 98 (C₃H₆O₂, 63%), respectively. The generation of such ions is only consistent with the cleavage of the C₇-C₁₁ and C₇-C₄ bonds, thereby strongly suggesting the presence of a primary hydroxyl group at C-14 or C-15, and a secondary hydroxyl group at C-3.

The existence of a secondary hydroxyl group in ring A at position 3 was reinforced by the following considerations: ¹H-NMR spectra of **1** and **2** depicted the presence of a non-allylic secondary carbinol group (m at

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4.6[†] and 5.15 ppm, respectively). These chemical shifts are comparable with those of compounds 15 and 3 (4.13 and 1.50 ppm), and different from those of compounds 16 and 17 (3.86 and 4.63 ppm).¹ The ¹³C NMR spectrum of 1 also agrees with these assignments (C-3: 81.01 ppm in 1 and 81.4 ppm in 15).

The complete structure of 1 was established by single crystal X-ray diffraction analysis. Crystals of 1 belong to the space group *p*2₁2₁2₁ with *a* = 18.44, *b* = 13.20, *c* = 6.51 Å, *Z* = 4. There are also eight water molecules in the unit cell. A total of 979 independent X-ray intensities with $\theta < 50^\circ$ were collected with Cu *K*_α radiation on a diffractometer. The structure was solved by direct methods and refined with isotropic thermal parameters (Table 1) by full-matrix least-squares to the conventional residual *R* = 0.12. The tetrol, given the same absolute configuration as Δ⁹⁽¹²⁾-capnellene-3β,8β,10α-triol 15,⁴ has similar conformation angles as one of the two independent molecules in the crystal structure of 15 (Fig. 1). There is one intramolecular hydrogen bond, O(4)–H...O(3), of 2.762(4) Å and seven other hydrogen bonds ranging from 2.73 to 2.81 Å. Thus all OH hydrogens are involved in hydrogen bonds.

Several other representatives of the capnellane class of sesquiterpenes are currently being investigated.

Table 1. Atomic coordinates ($\times 10^4$) and thermal parameters ($\times 10^4 \text{ \AA}^{-2}$) with e.s.d.'s in parentheses

	X	Y	Z	B
C(1)	3681(7)	5624(10)	28(2)	34(3)
C(2)	3066(6)	6400(9)	-6(2)	28(3)
C(3)	3491(6)	7402(9)	19(2)	25(2)
C(4)	4192(7)	7281(10)	-112(2)	31(3)
C(5)	4863(7)	7789(10)	-13(2)	31(3)
C(6)	5318(6)	6939(9)	91(2)	24(3)
C(7)	6144(7)	7094(11)	69(3)	43(3)
C(8)	6341(7)	6566(10)	-135(2)	33(3)
C(9)	5659(7)	5986(10)	-209(2)	29(2)
C(10)	5107(6)	5969(9)	-36(2)	26(3)
C(11)	4318(6)	6098(9)	-105(2)	22(2)
C(12)	5562(8)	5613(12)	-393(2)	43(3)
C(13)	4088(8)	7641(12)	333(2)	38(3)
C(14)	3472(7)	4593(10)	-57(2)	33(3)
C(15)	3824(7)	5516(10)	271(2)	35(3)
O(1)	3058(4)	8264(6)	-51(1)	36(2)
O(2)	6594(5)	7261(7)	-293(1)	38(2)
O(3)	5250(4)	5052(6)	88(1)	34(2)
O(4)	4082(4)	3902(6)	-38(1)	40(2)
O(W1)	1644(4)	421(6)	-223(1)	32(2)
O(W2)	2456(4)	8897(6)	413(1)	35(2)

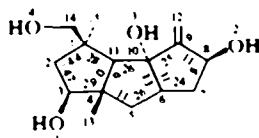
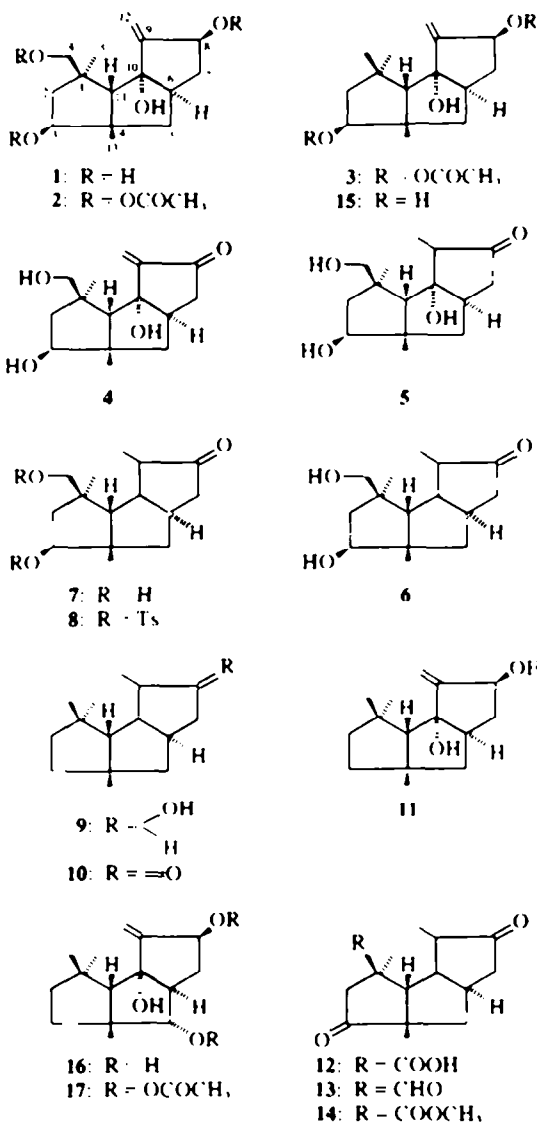


Fig. 1. Torsion angles for compound 1.

EXPERIMENTAL

M.p.s (Kofler) are uncorrected. All rotations and IR spectra were determined in chloroform solution. CD spectra were measured in MeOH. All NMR spectra were recorded (CDCl₃), unless otherwise mentioned, with TMS as internal standard) on a Varian T-60 or XL-100 spectrometer; all chemical shifts are reported in δ values. ¹³C NMR spectra were recorded using a

[†]This spectrum was recorded in 1:1 pyridine *d*₅-D₂O solution.



Varian XL 100 spectrometer operating at 25.5 MHz. Mass spectra (direct inlet system) were obtained by Mr. R. Ross with an AEI MS-9 or an Atlas CH-4 spectrometer and all peaks of relative intensity greater than 5% are reported. Mass spectral high resolution measurements were made by Miss Annemarie Wegmann using a Varian MAT 711 spectrometer. All GC analyses were carried out at oven temp. of 150–200° using a Hewlett-Packard 402 high efficiency gas chromatograph equipped with all-glass U-tube columns packed with 3% OV25, OV17 or OV3 stationary phases coated over Gas-Chrom Q (100–120 mesh). All TLC was performed using Merck silica gel F₂₅₄ compounds were visualized with ceric sulfate 2% in H₂SO₄2N. All compounds were found to be homogeneous by TLC and GC.

The general procedure for the isolation of the sesquiterpene of *Capnella imbricata* has been described.¹

Δ⁹⁽¹²⁾-Capnellene-3β,8β,10α,14-tetrol 1: m.p. 192–5°; $[\alpha]_D^{20} + 70^\circ$ (MeOH, *c* = 0.32) IR 3400 cm⁻¹; NMR (pyridine *d*₅-D₂O) 1.42,

1.68 (s, 3H each CH₃-C-), 3.82 (dd, 2H, CH₂OH), 4.60 (m, 1H,

CHOH), 5.33 (m, 1H, =C-CHO), 5.40 (m, 2H, CH=); MS: *m/e* 250 (5%, M-H₂O), 232(6), 219(18), 211(4), 202(17), 201(12), 199(10), 198(40), 149(27), 139(20), 131(13), 125(40), 123(17), 121(22), 112(37), 108(57), 107(100), 96(25), 95(43), 91(20), 81(17), 79(20), 69(17), 55(30). ¹³C NMR (TMS, MeOH): C-1(44.10), C-2(46.66), C-3(81.01), C-4(52.42), C-5(45.95), C-6(50.10), C-7(38.44), C-

8(73.84), C-9(161.90), C-10(89.11), C-11(63.54), C-12(109.09), C-13(24.80), C-14(73.84), C-15(21.96). On acetylation compound 1 furnished the triacetate 2: m.p. 122°; IR 1740 cm⁻¹; NMR: 0.95, 1.35 (s, 3H each, CH₃-C), 2.05 (s, 6H, 2x CH₃COO), 2.10 (s, 3H, CH₃COO), 4.10 (dd, 2H, CH₂OAc), 5.15 (m, 1H, CHOAc), 5.40 (bs, 2H, CH₂=), 5.83 (m, 1H, =C-CHOAc); MS: *m/e* 376 (1%, M-H₂O), 334 (13, M-CH₃COOH), 292(3), 274(27), 261(5), 214(13), 201(10), 181(7), 167(21), 121(4), 120(30), 108(55), 107(88), 94(49), 43(100).

Δ^9 -*Capnellene-3 β ,10 α ,14-triol-8-one* 4. To compound 1 (100 mg) dissolved in warm chloroform (200 ml) was added 1 g of active manganese dioxide, and the mixture stirred at room temp. in the dark overnight. After filtration and evaporation of the solvent, the residue was purified by preparative TLC to give oily 4: IR: 1700 cm⁻¹ (C=O); CD: [θ]₅₈₉ + 375, [θ]₅₁₇ - 3718, [θ]₂₄₀ + 8880; NMR (CDCl₃, acetone d₆): 0.90, 1.42 (s, 3H each, CH₃-C), 3.30 (dd, 2H, CH₂OH), 4.26 (m, 1H, CHOAc), 5.58, 6.16 (s, 1H each, CH₂=C); MS: *m/e* 266 (2%, M⁺), 248(3), 149(9), 143(5), 101(5), 91(6), 83(6), 81(3), 74(4), 71(5), 69(5), 59(55), 58(11), 57(10), 56(5), 55(10), 43(100).

On acetylation compound 4 furnished a diacetate: oil, IR: 1715 cm⁻¹ (C=O); NMR: 0.80, 1.40 (s, 3H each, CH₃-C), 2.03, 2.10 (s, 3H each, CH₃COO), 4.28 (dd, J = 11, 14 Hz, 2H, CH₂OAc), 5.13 (m, 1H, CHOAc), 5.50, 6.20 (s, 1H each, CH₂=C).

Δ^9 -*Capnellene-3 β ,14-diol-8-one* 6. To a soln of Li (100–150 mg) in 100 ml of liquid ammonia was added a soln of 4 (100 mg) in tetrahydrofuran (25–40 ml). After 30 min., solid ammonium chloride was slowly added until the blue soln decolorized. The ammonia was allowed to evaporate; water (50 ml) and ether (100 ml) were added and the mixture stirred vigorously for 10 min. After separation and drying over Na₂SO₄, the ether layer was evaporated to an oily residue. Crude 5 was dissolved in methanol (15 ml) to which five drops of 10% KOH were added, and the mixture stirred at room temp. for 6 h. The soln was then poured into equal volume of water and extracted twice with ethyl acetate (2 x 50 ml), washed with water, dried over Na₂SO₄, and evaporated to a gum which was purified by preparative TLC, to yield oily 6: IR: 1715 cm⁻¹ (C=O); NMR: 0.86, 1.10 (s, 3H each, CH₃-C), 1.66 (d, J = 1.5 Hz, 3H, CH₃-C=C), 3.50 (m, 2H, CH₂OH), 4.0 (m, 1H, CHOAc); MS: *m/e* 250 (23%, M⁺), 233(4), 232(20), 202(22), 179(12), 178(17), 175(11), 174(12), 161(17), 160(15), 159(11), 150(18), 149(20), 135(15), 133(14), 111(20), 109(24), 107(19), 105(22), ... 43(100).

Conversion of Δ^9 -capnellene-3 β ,14-diol-8-one 6 to capnellan-8-one 10. Li:NH₃ reduction of 6 (see above for experimental procedure) furnished oily capnellan-3 β ,14-diol-8-one 7: IR: 1720 cm⁻¹; MS: *m/e* 252.17384 (17%, M⁺ calculated for C₂₀H₃₀O₂: 252.17253) 234(9), 204(100), 177(39), 161(15), 135(15), 133(20), 121(24), 119(15), 109(42), 101(13), 94(40), 93(41), 91(29), 83(28), 79(31), 77(21), ...

On tosylation (*p*-TsCl/pyridine) compound 7 furnished a di-tosylate 8, which was reduced directly with LiAlH₄ in THF under reflux for 6 h to provide 9, which was oxidized with Jones reagent to 10 identical (GC, MS and IR) with the ketone derived from Δ^9 -*capnellene-8 β ,10 α -diol 15*.¹

Jones oxidation of 7. To a solution of the keto-diol 7 (5 mg) in acetone (5 ml) was added Jones reagent dropwise at room temp. for 15 min. The mixture was poured into water, extracted with ethyl acetate, washed with water, dried over Na₂SO₄, and evaporated to a gum. The analysis of this gum by TLC depicted one broad spot typical of a free carboxylic acid. To the gum were

added methanol (40 ml) and HCl (10N, 1 ml). The mixture was refluxed for 24 h, poured into ice and extracted with ethyl acetate. Preparative TLC furnished the more mobile diketo-ester 14 and the less mobile diketo-acid 12.

3,8-Dioxocapnellan-14-oic acid 12. IR: 1710–1730 cm⁻¹ (C=O); MS: *m/e* 264 (5%, M⁺), 151(100), 128(10), 114(70), 109(17), 107(25), 95(15), 94(14), 93(20), 91(9), 86(46), 81(7), 79(14), 77(8), 69(10), ...

Methyl 3,8-dioxocapnellan-14-oate 14. IR: 1735 cm⁻¹ (C=O); MS: *m/e* 278.15039 (M⁺, calculated for C₂₂H₃₂O₄: 278.15179), 128.04734 (C₈H₈O₃), 107(11), 100(24), 73(10), 69(17), 61(26), 45(27), 43(33).

The decarboxylation of 12 with *p*-TsOH and 2N HCl was attempted but in every case 12 was recovered unchanged.

Fetizon oxidation² of the keto-diol 7. A mixture of 7 (5 mg), Fetizon reagent (300 mg) and benzene (50 ml) was heated under reflux for 7 h. A portion (10 ml) of the benzene was distilled, but the remaining solution, when checked by TLC, depicted much unreacted starting material. The mixture was then refluxed for additional 72 h during which period almost all benzene was distilled. Preparative TLC of the residue furnished the oily 3,8-dioxo-capnellan-14-al 13: IR: 1725 cm⁻¹ (C=O); MS: *m/e* 248.14102 (6%, M⁺, calculated for C₁₈H₂₆O₂: 248.14123), 204(22), 151(31), 133(10), 108(28), 107(100), 98(63), 95(22), 94(49), 93(29), 91(18), 79(23), 77(15), 69(53).

This aldehyde could not be deformylated by heating with *p*-TsOH (in benzene) or aq KOH (in methanol).

Gas chromatographic retention times[†] relative to compound 1, of compounds 7, 10, 12 and 14

Compound No.	Relative retention time
1	1.00
4	1.10
10	0.20
12	0.65
14	0.41

[†]Over OV-3 3% on gas chrom Q (100–120), oven temp. 190° det. 290 inj. 260.

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